

IN VITRO ANTHELMINTIC ACTIVITY OF GOAT BILE CHEMICAL SUBSTANCES AGAINST Haemonchus contortus



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Abstract: Extracts of goat bile Chemicals substances were investigated for anthelmintic activity against *Haemonchus concortus*. Various concentrations (50, 100 and 200 mg/ml) of each extract of the bile were tested using the egg hatch assay and larval development assay. All extract tested showed 100% mortality at the end of 180 min of the test duration. While 0.25 mg/ml of albendazole the positive control showed 100% mortality after 120 min. The results of the egg hatch inhibition assay showed that at a concentration of 200 mg/ml, 65% of the eggs were inhibited from hatching by the bile extract. Albendazole inhibited 98% of the eggs from hatching at 0.25 mg/ml. Goat bile extract has potential as anthelmintic from nature, that can be explore in the search for natural remedy for parasitic affections.

Keywords: Albendazole, anthelmintic activity, egg hatching, goat bile, *Haemonchus concortus*

Introduction

Helminthiasis which is caused by the helminthes infection is proved to be a major constrain in the livestock production all around the globe. The parasitic disease affects the health status of a large fraction of human population as well as animals (Maity et al., 1998). In developing countries, they pose a large threat to public health and contribute to the prevalence of malnutrition, anemia, eosinophilia and pneumonia (Bundy, 2004). Chemical control of helminthes coupled with improved management has been the important worm control strategy throughout the world (Cole, 1997: Geert & Dorny, 1995). However, development of resistance in helminthes against conventional anthelmintics is a foremost problem in the treatment of helminthes diseases (Tagbota et al., 1994). Henceforth it is important to look for alternative strategies against gastrointestinal nematodes, which have led to the proposal of screening medicinal plants for their anthelmintic activity (Yoganandam et al., 2010).

Literature surveyed reveals that goat bile can be used to treat various kinds of gastrointestinal problems. Goat bile primarily consists of water, salts, cholesterol and a fatty substance (Barrett, 2012; Guyton and Hall, 2011). One of its main roles is to help the animal absorb vitamins from food that has been ingested, Goat biles were believed to be effective in treating optic atrophy, including acute hemorrhagic conjunctivitis, marginal suppurative blepharitis, and epiphora (Tao, 1955). Goat bile was also used to treat temporary blindness following a life-threatening illness (which was most likely smallpox), and eye injury from foreign bodies (ref). These biles were believed to be effective in ameliorating various infectious skin diseases, chancre in children (most likely impetigo), and constipation (ref). When compounded with pig pancreas and asarum herb, these biles were also prescribed as a facial lotion for minimizing chloasma in pregnant women and to derma base freckles (Li, 1957).

Also, when goat bile was decocted thrice with ox bile and wines the resulting liquor was believed to be effective in reversing any olive discoloration of the skin from itchy dermatitis, which was most likely secondary to tinea versicolor (Jiangsu, 1977). Therefore, the aim of this work is to evaluate the anthelminthic potentials of goat bile on *Heamonchus contortus*.

Materials and Methods

Sample collection

The one litre of goat bile was obtained from abattoir in Randan Kano, Zaira, Kaduna state of Nigeria in the month of

in July 2018. The goat bile sample was preserved with Methanol (40%) until needed for extraction.

Heamonchus contortus lavar

Adult *H. contortus* were recovered from the abomasums of freshly slaughtered goat in a local abattoir in Randan Kano, Zaria in the month of July, 2018. Which were confirmed by a Parasitologist at the Department of Parasitology, Faculty of Veterinary, Ahmadu Bello University, Zaria, Nigeria. All female *H. contortus* were morphologically selected and macerated to liberate the eggs. Distilled water was added and the eggs were successively sieved using Baermann funnel.

Extraction of Goat Bile

The bile (1 Litre) containing methanol (40%) was acidified with concentrated tetraoxosulphate (VI) acid (H₂SO₄), until the bile mixture was acidic to litmus paper. The observed precipitate (**BP-A**) was filtered. The filtrate was extracted with chloroform and concentrated under vacuum using a *rota vapor* at 40°C, to give chloroform bile extract (**CBE-1**). The aqueous layer was basified with NaOH until it was basic to litmus paper, the observed precipitate (**BP-B**) was filtered and the resulting aqueous layer extracted with chloroform and concentrated as earlier mention to give a second fraction (**CBE-2**). The precipitate BP-A and BP-B were suspended in ethylacetate and concentrated to remove any traces of water. *Biological studies*

Larval motility test (LMT)

Lava mortality test was conducted following the technique describe by Fernandez et al. (2009). Briefly; a total of about 368 lava parasites were used in the study. Three concentrations were employed for each extract. Ten worms were exposed in triplicate to each of the following treatments in separate Petri dishes at room temperature (25 - 30°C). There were 5 groups as follows; CBE-1, CBE-2, BP-A, albendazole (positive control) and water (negative control). The motility of worms was observed and mobile worms were counted at different time intervals till 7 h post treatment. Worms not showing any motility were picked out and kept in lukewarm phosphate buffer saline (PBS) for 10 min and, in case of revival in motility, the observed worms were counted as alive; otherwise, they were counted as dead. The Mortality rate for each concentration of the compound was determined using the ratio;

Total No.of initial larvae in wells – total No.of dead larvae in well Total number of initial larvae in wells x 100

Egg hatch inhibition assay (EHIA)

Freshly collected adult female H. contortus were picked. crushed and sieved to obtain the eggs, which were then triturated in PBS. The suspensions were centrifuged for 2 min at 300 rpm and the sediment was retained. This sediment was resuspended in saturated solution of NaCl to form a convex meniscus above the test tube. After putting a coverslip above the test tube, the samples were centrifuged again. Coverslip was carefully removed and eggs were washed into another test tube. This solution was then centrifuged and eggs were collected from sediment. Eggs were washed thrice with distilled water and adjusted to a concentration of 100 - 200eggs/mL, using the McMaster technique (Soul, 1982). Egg Hatch Inhibition Assay was performed following the technique of Coles et al. (1992). Approximately, 100 eggs in $200\,\mu$ l of water were pipetted into each well of a 48-well microtiter plate. To each of the test wells, $200 \,\mu$ l of each bile extract at different concentrations (50, 100 and 200 mg/ml) was added to a final volume of $400 \,\mu$ l per well. Similarly, $200\,\mu$ l of albendazole (standard drug) at $0.25\,\text{mg/ml}$ concentration and distilled water were used as positive and negative controls, respectively. Each test was carried out in three replicates and the results were expressed as mean \pm SD. The plates were incubated in a humidified incubator at 37°C x 100

 Table 1: Percentage mortality of Haemonchus contortus

for 48 h. Thereafter, a drop of Lugol's solution was added to stop further hatching. All unhatched eggs and larvae (L1) in each well were counted. The percent inhibition of egg hatching was calculated using the formula below (Coles et al., 1992). The percentage of hatched eggs was calculated using the ratio;

 $\frac{Total initial No. of eggs in wells - hatched eggs in well}{Total number of initial eggs in wells} x 100$

Results and Discussion

This study indicated that the bile extracts produced a relatively comparable anthelmintic activity with the conventional anthelmintic, albendazole. The activity increased with concentration and time. After 3 hours of exposure of Lavar of *H. contortus* to different concentrations of the bile extracts a dose-dependent increase in motility was observed for bile extracts (Table 1). At highest concentration (200 mg/mL), the bile extracts produced mortality of the larva *H. contorts* ranging 9 and 100% after 3 h exposure to the extracts (Table 1). Albendazole, on the other hand killed the parasites in a time-dependent manner and all the lavar worms were dead at a concentration of 0.25 mg/mL within 3 h of exposure.

Fractions	Concentration (mg/ml)	Mortality (Percentage ± SD)					
		30 min	60 min	90 min	120 min	150 min	180 min
CBE-1	200.0	12.7±3.35	63.7±2.65	87.7±5.65	100	100	100
	100.0	9.4±3.30	17.33±3.34	36.7±4.65	100	100	100
	50.0.0	0.00	11.7±6.65	13.0 ±3.0	46.0 ± 4.00	85.4 ± 5.30	100
CBE-2	200.0	18.0 ± 2.00	40.4 ± 1.70	88.4±2.30	100	100	100
	100.0	14.7 ± 5.85	35.0 ± 8.65	59.7±1.63	88.7 ± 2.00	100	100
	50.0	$11.0{\pm}1.00$	16.4 ± 7.00	36.7±4.65	55.7±4.35	65.7±5.25	100.0
BP-A	200.0	23.0 ± 4.70	50.0 ± 2.00	82.0 ± 2.00	$92.0{\pm}2.00$	100	100.0
	100.0	17.4 ± 2.70	44.0 ± 2.00	61.33 ± 5.60	87.7±2.60	90.0 ± 2.00	100.0
	50.0	10.0 ± 4.00	$24.4{\pm}15.1$	34.7±3.35	50.0 ± 2.00	80.0 ± 2.00	100.0
Albendazole	0.25	50	80	90.0	100.0	100.0	100.0
Water	1 ml	0.0	0.0	0.0	0.0	0.0	0.0
	0.0 = No mortality						

The result of egg hatch inhibition assay at graded concentration of bile extracts of is shown in Table 2. The result indicated that all the extracts produced a relatively comparable egg hatching inhibitory effect with albendazole. At a maximum concentration of 200 mg/ml CBE-1, CBE-2 and BPE-A induce egg hatch inhibition of 47, 59 and 64% while albendazole induced 98% egg hatching inhibitory at 0.25 mg/ml. Table 2.

 Table 2: Result of effect of the test samples at varying concentrations on egg hatching assay of H. contortus

Samples	Conc. (mg/ml)	Percentage inhibition
CBE-1	50	0.00
	100	26.0±1.80
	200	47.0±10.35
CBE-2	50	0.00
	100	22.2±1.65
	200	59.6±6.5
BPE-A	50	0.00
	100	47.7±1.70
	200	64.5±7.30
Albendazole	0.25	98±2.30
Water	0.00	0.00

In vitro tests to evaluate the inhibition of egg hatching and larval motility are widely used in veterinary parasitology to the prospecting of novel anthelmintic agents (Costa et al., 2002; Vasconcelos et al., 2007). The advantage of these assays is that compounds or materials to be tested are in direct contact with the different life-cycle stages of the parasite. Effective anthelmintic agents should inhibit worm egg hatching and larval motility by more than 90% and when inhibiting 80 - 90% should be considered moderately effective. Thus, our in vitro results obtained with the goat bile extract against H. contortus eggs and larvae, particularly in the higher dilutions, allow us to classify the tested extract as moderately effective (Ferreira et al., 2013). The bile extract showed a statistically significant anthelmintic activity against the eggs and larvae life-cycle stages of the parasite. This finding is significant since, theoretically, it reduces the chances of the occurrence of resistance of the parasite when using the extract in clinical practices due to anthelmintic action on different phases of worm development (Hounzangbe-Adote et al., 2005). Studies have suggested that the mechanism of inhibition of egg hatching and larval development of different parasites is related to the inhibition of cell division and, consequently, to the formation and development of vital structures of the parasite (Gallardo et al., 1998). Treatment of nematode infections using conventional

anthelmintic drugs resulted in about 294 million dollars of veterinary market revenue in 2004 (Molento et al., 2004). The consequences of this situation go beyond the rising costs of livestock management. In addition to the problem of resistance discussed above, there is no clear evidence that synthetic anthelmintics leave no residues in meat that would pose potential public health hazards (Rodrigues et al., 2007). Therefore, the identification of novel promising anthelmintic compound from bile extracts may contribute for the development of therapeutic products that could be more cost effective, safer, and more accessible and provide a lower risk of resistance than the conventional therapeutic arsenal currently employed. The in vitro tests using free living stages of parasitic nematodes offer a means of evaluating the anthelmintic activity of new compounds (Asase et al., 2005). In vitro techniques are preferred to in vivo methods due to their low cost, simplicity, and rapid turnover (Markus and Ernst. 2005). The bile chemical substance is documented to possess medicinal properties in ethnobotanical surveys conducted by ethnobotanists in traditional system of medicine (Wang and Carey, 2014). The present study showed 100% efficacy of the bile chemical substance extract against the parasite at the concentration of 200 mg/ml which was the highest efficacy value was comparable to the standard anthelmintic, albendazole. The egg hatching inhibitory effect of bile chemical substance extract was 64.5% at the concentration of 200 mg/ml. This is evident from the current study, which showed 95% of mortality of lavar parasites of H. contortus at a concentration of 200 mg/ml in all extracts of the goat bile chemical substance. The egg hatching inhibitory effect was highest (64%) at a concentration of 200 mg/ml for the BPE-A extracts of the goat bile chemical substance. Increment on the concentration of the extracts of the goat bile chemical substance resulted in increased inhibition of egg hatching indicating dose-dependent activity. Chemical substances like alkaloid and tannin have been reported to responsible for anthelmintic activity. The observed anthelmintic activity of plant extracts in the present study may be due to these chemical substances being present in the bile chemical substance (Debella, 2002; Da Silva, 2008).

Finally, the *in vitro* methods provide a means to screen rapidly for potential anthelmintic activities of different plant extracts. Due to drug biotransformation, interaction with food materials, and absorption variations, the results obtained by the *in vitro* method could not be extrapolated to *in vivo* activity. Therefore, results should be ascertained by *in vivo* evaluation. In conclusion, the current study showed that extracts of the bile chemical substance a promising *in vitro* anthelmintic activity against lavar and oval stages of *H. contortus.* Fractionation of the each extracts o of the bile chemical substance and further anthelmintic efficacy studies involving other parasite development stages are warranted.

Conflict of Interest

Authors declare that there is no conflict of interest reported in this work.

References

- Asase A, Oteng-Yeboah AA, Odamtten GT & Simmonds MSJ 2005. Ethnobotanical study of some Ghanaian antimalarial plants. *Journal of Ethnopharmacology*, 99(2): 1229–1221. doi: 10.1016/j.jep.2005.02.020.
- Barrett Kim E 2012. Ganong's Review of Medical Physiology (24th ed.). New York: McGraw-Hill Medical, p. 512. <u>ISBN 978-0-07-178003-2</u>.
- Bundy DA 1994. Immunoepidemiology of intestinal helminthic infection: The global burden of intestinal nematode disease. *Trans. Royal Soc. Trop. Med. Hyg.*, 8: 259-61.

- Coles GC, Bauer C, Borgsteede FHM *et al.* 1992. World Association for the Advancement of Veterinary Parasitology (W.A.A.V.P.) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. *Veterinary Parasitology.* 44(1-2): 35–44. doi: 10.1016/0304-4017(92)90141-U.
- Coles GC. Nematode control practices and anthelmintic resistance on British sheep farms. *Vet Rec.*, 141: 91-93. 4.
- Costa CTC, Morais SM, Bevilaqua CML, Souza MMC & Leite FKA 2002. Ovicidal effect of *Mangifera indica* L. seeds extracts on *Haemonchus contortus*. *Brazilian J*. *Veterinary Parasitol.*, 11: 57–60.
- Da Silva VC, De Carvalho MG, Borba HR & Silva SLC 2008. Anthelmintic activity of flavonoids isolated from roots of Andira anthelmia (Leguminosae). Revista Brasileira de Farmacognosia, 18(4): 573–576. doi: 10.1590/S0102-695X2008000400013.
- Debella A 2002. *Manual for Phytochemical Screening of Medicinal Plants*. Addis Ababa, Ethiopia: Ethiopian Health and Nutrition Research Institute.
- Gallardo T, Aragón R, Tormo JR, Blázquez MA, Zafra-Polo MC & Cortes D 1998. Acetogenins from Annona glabra seeds. *Phytochemistry*, 47: 811–816.
- Geert S & Dorny P 1995. Anthelmintic resistance in helminthes of animals of man in the tropics. Bulletindes-Seances, Academic Royaledes-Sciencesd. *Dutre Mer.*, 3: 401-423.
- Guyton & Hall 2011. Textbook of Medical Physiology. US Saunders Elsevier, p. 784. <u>ISBN 978-1-4160-4574-8</u>.
- Hounzangbe-Adote, M.S., Paolini, V., Fouraste, I., Moutairou, K., Hoste, H., 2005. In vitro effects of four tropical plants on three life-cycle stages of the parasitic nematode Haemonchus contortus. Research in Veterinary Science 78, 155–160.
- Husaain A, Khan MN, Iqbal Z, Sajid MS & Khan MK 2011. Anthelmintic activity of *Trianthema partulacastrum* L. and *Musa paradisiacal* L. Aainst Gastrointestinal Nematodes of Sheep, Vet Parasitology.
- Jiangsu New Medical College 1977. Great Dictionary of Chinese Materia Medica. Shanghai: People's Publishing House of Shanghai [Google Scholar].
- Kavitha S & Manimekalai GA 2015. Study on properties of Cissus quadrangularis plant: A review. Int. J. Res. Appl. Nat. and Soc. Sci., 3(6): 15–18.
- Li SZ 1957. Compendium of Materia Medica (Ben Cao Gang Mu) Beijing: People's Health Publishing House, 1596, Reprinted [Google Scholar].
- Luseba D, Elgorashi EE, Ntloedibe DT & Van Staden J 2007. Antibacterial, anti-inflammatory and mutagenic effects of some medicinal plants used in South Africa for the treatment of wounds and retained placenta in livestock. *South Afri. J. Botany*, 73(3): 378–383. doi: 10.1016/j.sajb.2007.03.003.
- Maity TK, Mandal SC, Mukherjee PK, Saha K & Das J 1989. Studies on antiimflammatory effect of Cassia tora Leaf extract. *Phytotherapy Research*, 12(2): 221-223.
- Markus S & Ernst M 2005. *Medicinal Plants in Tropical Countries*. Rudigerstrasse, Germany: Georg Thieme Verlag.
- Martin RJ 1997. Modes of action of anthelmintic drugs. *The Veterinary Journal*, 154(1): 11–34. doi: 10.1016/S1090-0233(05)80005-X. [PubMed] [CrossRef] [Google Scholar]
- Molento MB, Tasca C, Gallo A, Ferreira M, Bononi R & Stecca E 2004. Método Famacha como parâmetro clínico individual de infecção por Haemonchus contortus em pequenos ruminantes. *Ciência Rural*, 34: 1139–1145.

- Rodrigues AB, Athayde AC, Rodrigues OG, Silva WW & Faria EB 2007. Evaluation of the efficacy of anthelmintics to control gastrointestinal nematodes in goats raised in the state of Paraiba. *Pesquisa Veterinaria Brasileira*, 27: 162–166.
- Soulsby EJL 1982. *Helminths, Arthropod and Protozoa of Domestic Animals*. London, UK: The English Language Book Society and Bailliere Tindall.
- Srinivasa MV & Jayochandran E 2006. Anthelmintic activity of 8- flouro-9-Substituted(1,3)-benzthazole(5, 1-B)1,2,3triazole of *Pheretima poshuma*. *Indian Drug*, 403(4): 343-347.
- Tao HJ 1955. Variorum of Shen Nung's Herbal Classic (Ben Cao Jing Ji Zhu) Shanghai: Shanghai Qunlian Publishing House, c. 492 CE, reprinted. [Google Scholar]
- Vasconcelos ALC, Bevilaqua CM, Morais SM, Maciel M, Costa CT, Macedo IT, Oliveira LM, Braga RR, Silva RA & Vieira LS 2007. Anthelmintic activity of Croton

zehntneri and Lippia sidoides essential oils. Veterinary Parasitology, 148: 288–294.

- Wang GX, Zhou Z, Jiang DX, et al. 2010. In vivo anthelmintic activity of five alkaloids from Macleaya microcarpa (Maxim) Fedde against Dactylogyrus intermedius in Carassius auratus. Veterinary Parasitology, 171(3-4):305–313. doi: 10.1016/j.vetpar.2010.03.032.
- Wang HQ & Carey MC 2014. Therapeutic uses of animal biles in traditional Chinese medicine: An ethnopharmacological, biophysical chemical and medicinal review. World J. Gastroenterol., 20(29): 9952–9975.
- Yoganandam GP, Ilango K, Kumar Sunil & Elumalai A 2010. In-vitro antioxidant activity of L. cylindrica seeds oil. J. Global Pharma. Techn., 2(3): 93-97.